# RESEARCH

# **Open Access**



# Assessing current and future areas of ecological suitability for *Lutzomyia shannoni* in North America

Sydney DeWinter<sup>1\*</sup>, Grace K. Nichol<sup>1</sup>, Christopher Fernandez-Prada<sup>2</sup>, Amy L. Greer<sup>3</sup>, J. Scott Weese<sup>4</sup> and Katie M. Clow<sup>1</sup>

# Abstract

**Background** In the Americas, sand flies of the *Lutzomyia* genus are the vectors of pathogens of human and animal health significance. *Lutzomyia shannoni* is suspected to transmit vesicular stomatitis virus, along with *Leishmania mexicana* and *Leishmania infantum* (causative agents of leishmaniases). Despite the suspected vector potential of *Lu. shannoni*, significant knowledge gaps remain, including how ongoing climate changes could facilitate their range expansion. The objectives of this study were to predict the current and future ecological suitability of regions across North America for *Lu. shannoni* and to identify variables driving ecological suitability.

**Methods** Occurrence records were obtained from the Global Biodiversity Information Facility, Disease Vectors Database, the National Museum of Natural History (Smithsonian Institution) and published literature on *Lu. shannoni* surveillance and capture. Historical climate data from 1991–2020, along with projection data for Shared Socioeco-nomic Pathways 2–4.5 and 3–7.0 were obtained. An additional terrestrial ecoregions layer was applied. The ecological niche model was created using maximum entropy (MaxEnt) algorithms to identify regions which currently are or may become ecologically suitable for *Lu. shannoni*.

**Results** Currently, regions in eastern, western and southern Mexico, along with the Midwest, southeastern and eastern regions of the USA are ecologically suitable for *Lu. shannoni*. In the future, ecological suitability for *Lu. shannoni* is expected to increase slightly in the northeastern regions of the USA and in Atlantic Canada, and to decrease in the southeastern reaches of Mexico. Degree-days below 0 °C (spring and autumn), precipitation as snow (summer and winter), terrestrial ecoregions, number of frost-free days (summer), Hargreaves climatic moisture deficit (summer), degree-days above 5 °C (autumn) and Hogg's climatic moisture index (summer) were all identified as predictors of ecological suitability.

**Conclusions** The findings from this study identified climate and environmental variables driving the ecological suitability of regions for *Lu. shannoni* and can be used to inform public health professionals of high-risk regions for exposure at present and into the future.

**Keywords** *Lutzomyia shannoni, Leishmania* spp., Vesicular stomatitis virus, Climate change, Range expansion, Ecological suitability, Ecological niche modeling

\*Correspondence: Sydney DeWinter sdewinte@uoguelph.ca Full list of author information is available at the end of the article



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/A.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

# Background

Phlebotomine sand flies, such as members of the Lutzo*myia* genus, are small (approx. 2 mm in size) pool feeders (i.e. blood-feeders that cut a hole in the skin and feed on the blood that pools from the wound) that are currently well-distributed in the Americas [1–10]. *Lutzomyia* spp. can be found in agricultural or rural areas of tropical and subtropical regions in countries such as the USA, Mexico, Guatemala, Belize, El Salvador, Nicaragua, Costa Rica, Ecuador, Colombia, Venezuela, Guyana, French Guiana, Suriname, Brazil, Argentina, Paraguay and Bolivia [1-4, 6, 8, 11]. Their life-cycle consists of four major stages: eggs, larvae, pupae and adults [1]. Female Lutzomyia spp. can live 2-6 weeks and undergo multiple gonotrophic cycles, and each cycle can produce anywhere from 30 to 70 eggs [1, 7, 8, 12, 13]. While adult sand flies consume plant sugars, female sand flies must also take multiple blood meals from mammals to produce eggs [1, 7, 8, 12, 13]. It is during these blood meals that *Lutzomyia* spp. females may ingest and then subsequently transmit-in a future meal-pathogens such as vesicular stomatitis virus (VSV) and Leishmania spp. to infective mammalian hosts [5, 7, 8, 14, 15].

VSV causes disease in horses, ruminants and other livestock species, including swine. Blister-like lesions form in the mouth, lips, ears, hooves and udders, causing discomfort and a significant reduction in productivity. Moreover, the lesions caused by VSV closely resemble those associated with foot and mouth disease virus and thus can result in pre-emptive quarantine and trade disruptions. VSV can also cause disease in humans [14, 15]. Leishmania spp. are the etiologic agent of leishmaniases in both humans and animals, namely dogs. Leishmaniasis is a global health issue that impacts approximately 2.5 million dogs and 12 million people, with over 700,000 new cases diagnosed annually [16, 17]. In humans, leishmaniasis can manifest as a visceral, cutaneous or mucocutaneous form, all of which are caused by different Leishmania spp. Visceral leishmaniasis (caused by Leishmania infantum and Leishmania donovani) is characterized by damage to the hosts' internal organs, while cutaneous leishmaniasis (caused by Leishmania mexicana, Leishmania braziliensis and Leishmania major) causes skin lesions [16, 17]. Mucocutaneous leishmaniasis (caused by L. braziliensis and Leishmania guyanensis) causes damage to the mucous membranes of the host [14-16]. In dogs, infection with L. infantum or L. braziliensis is a multisystemic disease in nature, involving almost all the hosts' body systems [16, 17, 19-24]. Common signs of infection include lymphadenopathy and dermatological abnormalities, with disease progression leading to renal failure and death [16, 17, 24]. Dogs also play a unique epidemiological role in *Leishmania* spp. transmission as they serve as reservoir hosts for *L. infantum* and *L. braziliensis*.

In North America, one of the northern-most sand flies, Lutzomyia shannoni, is a potential vector of VSV, along with Leishmania spp. Lutzomyia shannoni meets the requirements of a biological vector for the VSV, with vertical transmission having been demonstrated in laboratory studies and possibly in nature [14]. Further, laboratory studies have found Lu. shannoni to be capable of acquiring L. mexicana from infected animals and subsequently transmitting it during a subsequent blood meal [25, 26]. However, the relationship between the parasite and vector has not been confirmed in nature [25]. Lutzomyia shannoni has also been suspected to transmit L. infantum, as the former were reported in areas where L. infantum-infected dogs were reported, and L. infantum remains infective in Lu. shannoni following ingestion, although further confirmatory studies are lacking [27]. While current research suggests that Lu. shannoni demonstrate a lower infection rate compared to other sand flies, such as *Lu. longipalpis*, and infection has not been documented definitively in the USA, this sand fly remains a potential concern for domestic animal and human health as its range extends into more northern areas where other sand fly species are absent [27]. Lutzomyia shannoni has been found across 14 U.S. states, and as far north as the state of New Jersey [11, 15].

Over the past several decades, countries across the world have experienced climatic shifts due to anthropogenic carbon emissions and subsequent global warming [9, 10]. These climatic shifts have resulted in the expansion of ecological suitable areas for several vector species, such as ticks and mosquitoes [28–32]. It is unknown if sand flies such as *Lu. shannoni* may expand or shift their range under climate change and thus provide a mechanism for local transmission. This is particularly relevant as there is considerable international domestic animal movement, which could introduce VSV or *Leishmania* spp.

Ecological niche models (ENM) are statistical models that use environmental data to predict the distribution of a species (such as *Lu. shannoni*), based on ecological suitability [33–36]. These models can be utilized for a variety of reasons, such as when a species' distribution is not well-defined and surveillance data are lacking. Maximum entropy (MaxEnt) algorithms are a type of ENM that utilize presence-only data [37]. As *Lu. shannoni* is an under-studied *Lutzomyia* sp., these models can be used to understand both their current potential distribution based on ecological factors, as well as regions of future ecological suitability, under different climate change scenarios. Therefore, the objectives of this study were to (i) predict the current and future ecological suitability in North America for *Lu. shannoni*, and (ii) to identify variables impacting *Lu. shannoni* ecological suitability. As *Lu. shannoni* distribution is dependent on locally occurring ecological factors, such as precipitation, temperature and habitat availability (i.e. the presence of deciduous trees), such factors are expected to have considerable impact on ecological suitability both currently and into the future [15, 38, 39].

# Methods

#### Study area

The terrestrial regions of North and Central America (as far south as Mexico) were utilized for ecological niche model generation. This extent captured the current distribution of *Lu. shannoni*, and the northward regions of interest for range expansion.

## Data acquisition and preparation

Presence-only data, including coordinates and year of collection, for *Lu. shannoni* were obtained from the Global Biodiversity Information Facility (GBIF) (https://www.gbif.org), the Disease Vectors Database (discontinued) and the National Museum of Natural History (Smithsonian Institution) (https://naturalhistory.si.edu). Additional presence points were obtained from previous surveillance data reported in the literature [2–4, 6, 11, 40–52]. The distance between presence points was investigated in QGIS version 3.22.1 (https://qgis.org/en/site; 2024). To reduce artificial clustering and spatial bias correction, all duplicate presence points and points less than 3.5 km apart were removed from further analyses [53].

Climate variables derived from daily weather data, such as temperature (minimum, maximum, average), precipitation values, frost-free days and degree days, were obtained from ClimateNA (1991-2020) (https:// climatena.ca). Fifteen seasonal climate variables (60 variables total, one for each season) were downloaded at a resolution of 4 km<sup>2</sup>. An additional data layer, containing terrestrial ecoregions (level III), were obtained from the Commission for Environmental Cooperation (CEC; http://www.cec.org) (Table 1). Projection data were obtained from ClimateNA for emission scenarios based on socioeconomic predictions, known as Shared Socioeconomic Pathways (SSPs). The General Circulation Models (GCMs) for Shared Socioeconomic Pathways 2–4.5 and 3–7.0 (i.e. the 'middle of the road' and 'upper middle') were selected. SSP 2.4-5 assumes temperatures would rise by 2.7 °C by 2100, and SSP 3-7.0 assumes temperatures would rise by 3.6 °C by 2100 [54]. Data were projected into two 30-year periods: 2041-2070 and 2071–2100. Ecoregions remain stable over time; therefore, the same ecoregion layer was retained in both the base environmental layer and projection layer.

Data were imported into QGIS Version 3.22.1 to be rasterized. Data were geoprocessed to the same resolution and coordinate reference system (CRS84). Climate data at each presence point were extracted from QGIS and imported into RStudio (version 4.2.1; R Foundation for Statistical Computing, Vienna, Austria). A correlation matrix was created, and when highly correlated (>0.80), retained variables were chosen based on ecological importance for *Lu. shannoni*, based on previous findings [10]. In the instance where > 1 highly correlated variable had ecological relevance, all were retained for initial model iteration.

## Ecological niche model—current projection

The initial ecological niche model was based on historic data. Presence data and rasterized ecological data were imported into MaxEnt species distribution modeling software (version 3.4.4) [53]. A k-fold cross-validation run-type was used to examine the data, where k=4 [35, 37, 55–57]. Linear, guadratic and product feature classes were selected [35, 37, 55-57]. To prevent overfitting of the model, the regularization multiplier was increased [35, 37, 55–57] (Table 2). In previous research, the regularization multiplier has been changed from the default of 1.0, to values ranging from 1.5 to 3.0 [35, 37, 55–57]. A regularization multiplier of 1.0, 1.5 and 2.0 were applied, with the iteration returning the highest area under the receiver operating curve (ROC) (AUC) and lowest average omission rate being selected for projection layer application. The final model, with a regularization multiplier of 1.5, was chosen. An iterative approach was taken when building the ecological niche model, wherein the permutation importance of each variable was assessed. If a variable's permutation importance was 0%, it would be removed from the model, and variables included in the following iteration had an importance of > 0%. When highly correlated variables were included in the final model, the correlated variable with the higher permutation importance was retained, while the other was removed. The final model was comprised of variables with a permutation importance of>0%, and no highly correlated variables. Independent response curves for each environmental variable, along with a jackknife test of regularized training gain were generated.

#### Ecological niche model—future projections

Variables included in the final current ecological niche model were considered to have an impact on *Lu. shannoni*, and therefore carried forward when generating future projection maps. No retained variables were highly correlated.

Variable	Description	Included in final model?	
CEC terrestrial ecoregions level III	Ecological regions are areas of similarity in ecosystems and environmental resources Level III ecoregions are smaller ecological areas within larger ecoregions	Yes	
CMDsm	Hargreaves climatic moisture deficit (mm) [summer]	Yes	
CMIsm	Hogg's climate moisture index (mm) [summer]	Yes	
CMIsp	Hogg's climatic moisture index (mm) [spring]	No	
DD0at	Degree-days below 0 °C [autumn]	Yes	
DD0sm	Degree-days below 0 °C [summer]	No	
DD0sp	Degree-days below 0 °C [spring]	Yes	
NFFDsm	Number of frost-free days [summer]	Yes	
NFFDsp	Number of frost-free days [spring]	No	
NFFDwt	Number of frost-free days [winter]	No	
NFFDat	Number of frost-free days [autumn]	No	
PASat	Precipitation as snow [autumn]	No	
PASsm	Precipitation as snow [summer]	Yes	
PASsp	Precipitation as snow [spring]	No	
PASwt	Precipitation as snow [winter]	Yes	
PPTsm	Precipitation (mm) [summer]	No	
PPTsp	Precipitation (mm) [spring]	No	
PPTwt	Precipitation (mm) [winter]	No	
Taveat	Mean temperature (°C) [autumn]	No	
Tavesp	Mean temperature (°C) [spring]	No	
Tavesm	Mean temperature (°C) [summer]	No	
Tavewt	Mean temperature (°C) [winter]	No	
Tmaxwt	Mean maximum temperature (°C) [winter]	No	
Tmaxsp	Mean maximum temperature (°C) [spring]	No	
Tmaxat	Mean maximum temperature (°C) [autumn]	No	
DD5wt	Degree-days above 5 °C [winter]	No	
DD5at	Degree-days above 5 °C [autumn]	Yes	
DD5sp	Degree-days above 5 °C [spring]	No	
DD18at	Degree-days above 18 °C [autumn]	No	
DD18sp	Degree-days above 18 °C [spring]	No	
Tminat	Mean minimum temperature (°C) [autumn]	No	
Tminsp	Mean maximum temperature (°C) [spring]	No	

Table 1 Summary of environmental and bioclimatic variables investigated in Lutzomyia shannoni ecological niche model construction

CEC Commission for Environmental Cooperation

## Model evaluation

Model fit was evaluated using the mean AUC and the test omission rate of the minimum training presence.

**Table 2** Initial maximum entropy model parameter settings

 applied in ecological niche model construction to investigate
 ecological suitability for *Lutzomyia shannoni* in North America

Parameter setting	
quadratic, product	
bg	
cross-validation	

The model returning the highest AUC (i.e. closest to 100%) and lowest omission rate (i.e. closest to 0) was the best fitting model and therefore considered to be the final ecological niche model.

# Results

## Vector species records

Following the removal of duplications and rarefaction of coordinates, 80 presence points from 1991 to 2020 were eligible for inclusion. Species records were collected as far south as the states of Oaxaca and Veracruz, Mexico. The most northern records were collected from the U.S. states of Ohio, Maryland and New Jersey (Fig. 1).

#### **Ecological niche model**

Terrestrial ecoregions and 31 bioclimatic variables were included in initial model building [10] (Table 1). After removing all variables with a permutation importance of 0, no remaining variables were highly correlated. The final model predicting the ecological suitability of *Lu. shannoni* in North America included degree-days below 0 °C (autumn) (permutation importance of 73.6%), precipitation as snow (summer) (8.0%), precipitation as snow (winter) (5.3%), degree-days below 0 °C (spring) (4.7%), CEC terrestrial ecoregions level III (2.4%), number of frost-free days (summer) (2.3%), Hargreaves climatic moisture deficit (summer) (1.8%), degree-days above 5 °C (autumn) (1.2%) and Hogg's climatic moisture index (summer) (0.6%) (Fig. 2; Tables 2, 3).

The model had an AUC of 97.4%, with a standard deviation (SD) of  $\pm$  0.003. According to Fielding and Bell [55], this AUC value was indicative of a good model fit. Using the mean minimum training presence of the test data, we determined the omission rate to be 0.027 (Table 3). Based on the independent response curves of the ecological variables, suitability for Lu. shannoni increased when the Hogg's climatic moisture index in the summer, degreedays above 5 °C in the autumn and the number of frostfree days in the summer increased. Additionally, regions ecologically suitable for Lu. shannoni decreased when there were increases in the degree-days below 0 °C in the spring and autumn, Hargreaves climatic moisture deficit in the summer and precipitation as snow in the summer and winter. Some terrestrial ecoregions were associated with an increase in ecological suitability (i.e. predicted ecological suitability of>0.70), including the Mississippi valley loess plain; ridge and valley; southern coastal plain; Sierras of Guerrero and Oaxaca with conifer, oak and mixed forests; Chiapas highlands with conifer, oak and mixed forest; south Pacific hills with and piedmonts with low tropical deciduous forest; Gulf of Mexico coastal plain with wetlands and high tropical rain forest; hills with medium and high evergreen tropical forest; plain with low and medium deciduous tropical forest and hills with high and medium semi-evergreen tropical forest; Los Tuxtlas Sierra with high evergreen tropical forest;



Fig. 1 The geographic location of presence points for *Lutzomyia shannoni* (n = 80). Presence points were identified on the basis of human or machine observation, from 1991 to 2020. Map was constructed in QGIS (version 3.22.1)



Fig. 2 Independent response curves depicting the dependence of predicted ecological suitability for *Lutzomyia shannoni* on each modeled environmental variable. The red bands indicate the mean response, and the blue bands indicate the standard deviation. Graphs were constructed using the maximum entropy (MaxEnt) (version 3.4.4) algorithms [56]

Environmental variable	Permutation importance (%)	Area under the receiver operating characteristic curve	Mean test omission rate (minimum training presence)
CEC terrestrial ecoregions level III	2.4		
Hargreaves climatic moisture deficit (summer)	1.8		
Hogg's climatic moisture index (summer)	0.6		
Degree-days below 0 °C (spring)	4.7		
Degree-days below 0 °C (autumn)	73.6	97.4%±0.003	0.027
Degree-days above 5 °C (autumn)	1.2		
Number of frost-free days (summer)	2.3		
Precipitation as snow (winter)	5.3		
Precipitation as snow (summer)	8.0		

	Table 3	Summar	of maximum	entropy outputs f	or the final eco	logical niche model
--	---------	--------	------------	-------------------	------------------	---------------------

CEC Commission for Environmental Cooperation

and Jalisco/Nayarit hills and plains with medium semievergreen tropical forest . Results of the jackknife test of variable importance indicated that the variable with the highest independent gain was the CEC terrestrial ecoregions level III layer. Further, when this variable was omitted, the model gain decreased the most (Figs. 2,3).

#### Predicted current ecological suitability, 1991–2020

Regions predicted to be currently ecologically suitable for *Lu. shannoni* in Mexico included the majority of southern Mexico, including both eastern and western extents. Much of central and northern Mexico is not currently ecologically suitable for *Lu. shannoni*. In the USA, suitable regions were across the Midwest, southeastern and eastern states. The west coast of the USA was not ecologically suitable for *Lu. shannoni*. In Canada, the coastal region of the province of British Columbia was found to have low ecological suitability. Terrestrial ecoregions suitable for *Lu. shannoni*, from 1991 to 2020, were variable in their characteristics, but could be generally defined by their high humidity and forest cover, specifically, deciduous and tropical forest types (Fig. 4).

## Projected future ecological suitability, 2041-2070

*Shared socioeconomic pathway* 2–4.5 Based on this climate projection, there was a constriction of ecological suitability in southeastern Mexico and in the Midwest of the USA. Western Mexico and northeastern regions of the USA are forecasted to expand in their ecological suitability for *Lu. shannoni* (Fig. 5).

*Shared Socioeconomic Pathway* 3–7.0 With this climate projection, ecological suitability for *Lu. shannoni* decreased along the southeastern regions of Mexico, and slightly decreased in the Midwest of the USA. There was a slight increase in ecological suitability noted again in the

western regions of Mexico, as well as along the southeastern and northeastern regions of the USA (Fig. 5).

#### Projected future ecological suitability, 2071-2100

*Shared socioeconomic pathway* 2–4.5 The largest general expansion of ecological suitability was observed with this climate projection. Under SSP 2–4.5, constriction in southeastern Mexico and the Midwest of the USA was noted again. Regions such as western Mexico, northeastern USA, Atlantic Canada and coastal British Columbia were all forecasted to increase in terms of their ecological suitability for *Lu. shannoni* (Fig. 5).

*Shared socioeconomic pathway* 3–7.0 This projection forecasted considerable constriction across southern and southeastern Mexico, and again in the Midwest of the USA. Ecological suitability for *Lu. shannoni* was forecasted to increase in northeastern regions of the USA (Fig. 5).

## Discussion

The ongoing impacts of climate change on ecological niche shifts of many dipteran vectors have been well-documented [30, 31]. Many of these dipterans are capable of transmitting pathogens infective to humans and animals, and *Lutzomyia* spp. are no exception. Continuous investigation into the ecological niche of *Lutzomyia* spp., including *Lu. shannoni*, is important to gain



Fig. 3 Jackknife test of variable importance for *Lutzomyia shannoni* ecological niche model. This test determines the regularized training gain of each variable in the final model, demonstrating which variables have the greatest impact on model gain when in isolation (indicated in blue), or when omitted from the model (indicated in teal). Graph was constructed using the maximum entropy (MaxEnt) (version 3.4.4) algorithms [56]



Fig. 4 Current ecological suitability model for *Lutzomyia shannoni*, using climate and ecoregion data from 1991–2020. Warm colors indicate areas of high suitability, whereas cool colors are indicative of areas with low suitability. Map was constructed using the maximum entropy (MaxEnt) (version 3.4.4) algorithms [56]

an understanding of which regions are currently suitable, and which could become suitable under changing climate conditions. In this study, we elucidated information on the current and future ecological suitability of North America for Lu. shannoni, a potential vector of VSV, L. mexicana and L. infantum [2, 7, 15, 20]. In the model utilized, ecological suitability was greatly impacted by degree-days below 0 °C (autumn), precipitation as snow (summer and winter), degree-days below 0 °C (autumn) and, to a lesser extent, terrestrial ecoregions, number of frost-free days (summer), Hargreaves climatic moisture deficit (summer), degree-days above 5 °C (autumn) and Hogg's climatic moisture index (summer). Under the investigated climate scenarios, there were slight shifts in suitability across southern Mexico, and additional shifts across the eastern USA, coastal British Columbia and Atlantic Canada.

Degree-days below 0 °C (autumn) and terrestrial ecoregions had the highest permutation importance and greatest gain in the model, respectively. Therefore, it can be inferred that shifts in ecological suitability for *Lu. shannoni* are largely driven by these two variables. Degreedays are the number of days when the temperature is above or below a fixed reference value, typically with reference to vector development [58]. This parameter has been long identified as a major driver for vectors, namely due to its impact on vector activity and development [58]. In the context of this research, fewer degree-days below 0 °C in the autumn and spring were positively associated with ecological suitability. Further, ecological suitability increased with increasing number of degree-days above 5 °C in the autumn. Indeed, warmer transitional seasons (spring and autumn) would provide more developmentally favorable conditions for Lu. shannoni [5, 8-10, 59, 60]. Degree-days are a temperature-dependent variable, and the importance of temperature for *Lutzomyia* spp. development and activity has been well documented. For example, previous studies have reported that temperatures above 15 °C are ideal for Lutzomyia spp., but other studies have reported that minimum temperatures need to remain above 10 °C for at least 3 months for Lutzo*myia* spp. to become established [7, 10, 61].

Terrestrial ecoregions are ecological regions with general similarities between their ecosystems, including the environmental resources they possess [62]. The CEC terrestrial ecoregions (level III) are smaller ecological areas nested into larger ecoregions [63, 64], allowing for the elucidation of more specific habitat information.



Fig. 5 Future ecological suitability model for *Lutzomyia shannoni*, from 2041 to 2070, and from 2071 to 2100, under shared socioeconomic pathways (SSP) 2–4.5 and 3–7.0. Warm colors indicate areas of high suitability, and cool colors are indicative of areas with low suitability. Map was constructed using the maximum entropy (MaxEnt) (version 3.4.4) algorithms [56]

Information on the ecoregion requirements for Lu. shannoni is fundamental to determining their ecological niche. Ecoregions provide distinct boundaries, allowing for extrapolation from presence points to whole ecoregions. Additionally, the common characteristics defining ecoregions provide additional ecological information regarding the niche of Lu. shannoni. In this study, numerous ecoregions were identified as ecologically suitable for Lu. shannoni, being defined by generally warm, temperate climates with a mix of deciduous, evergreen and tropical forests. Specifically, it is known that adult Lu. shannoni can be found in tree holes of deciduous trees [15, 38]. Therefore, ecoregions with this vegetation present likely serve as refugia sites. Despite the clear importance of ecoregions when elucidating the ecological niche of these sand flies, they have been rarely incorporated into ecological niche models of other Lutzomyia spp. [5, 9].

Due to the sensitivity of *Lu. shannoni* to temperature, it is known that frost and snow negatively impact their development [65, 66]. From the model, an increase in snow or frost was associated with a decrease in ecological suitability. Both snow and frost can only form at low temperatures, specifically those of less than 0 °C [65, 66]. As stated above, it is known *Lu. shannoni* require temperatures consistently higher than 0 °C for establishment to occur [7, 10, 61]. While the presence of snow and frost has negative impacts on *Lu. shannoni*, these factors are also an extension of temperature, specifically low ones, that are not conducive to their development, activity and potential establishment. Further, *Lu. shannoni* are small (2 mm) and considered to be weak fliers [1–10]. Therefore, snow presents a physical issue for *Lu. shannoni*, which are known to be sensitive to both wind and heavy precipitation.

Hargreaves climatic moisture deficit and, to a lesser extent, Hogg's climatic moisture index (in the summer) were also found to be important when determining the ecological niche of Lu. shannoni, albeit to a lesser extent than degree-days and terrestrial ecoregions. Hargreaves climatic moisture deficit is calculated as potential evapotranspiration against actual evapotranspiration [67]. This measurement is strongly correlated with the distribution of vegetation in a landscape, and the moisture deficit accumulates over the season. The model generated in this study reported an inverse relationship between the moisture deficit and ecological suitability, with an increase in the moisture deficit being associated with a decrease in ecological suitability for Lu. shannoni [67]. Hogg's climatic moisture index is an indicator of drought, with positive values being indicative of wet or moist conditions able to sustain a close-canopy forest [64]. In

this ecological niche model, a higher moisture index was associated with an increase in ecological suitability. The importance of precipitation and moisture in the ecological niche of *Lutzomyia* spp. sand flies is well established [5, 8-10]. It is known that environmental moisture supports *Lutzomyia* spp. development, along with the creation of refugia [5, 8-10].

Based on the results from the presence records that were obtained, Lu. shannoni is currently found throughout southern and central Mexico and into central and eastern USA. The results from our ecological niche model align well with the known distribution of Lu. shannoni but they did identify some regions that are currently ecologically suitable, but from which no records have been recorded. These regions include coastal British Columbia and some parts of western Mexico and northeastern USA. Some regions of constriction and expansion were noted in the future projections. Generally, model projections predicted a decrease in ecologically suitable habitats across southeastern regions of Mexico and the Midwest regions of the USA, with many regions in Mexico and the USA forecasted to remain ecologically suitable. In Canada, regions such as coastal British Columbia and some reaches of the Maritimes were forecasted to become suitable, albeit only slightly.

It is important to note that even if areas are deemed currently ecologically suitable, Lu. shannoni require a dispersal mechanism to facilitate range shifts (e.g. human intervention, extreme weather events, among others). The data utilized in ecological niche models are not suitable to investigate specific dispersal mechanisms. For example, previous research has implicated windborne incursions of other vectors, such as biting midges (Culicoides spp.) and black flies (Simulium spp.), across 600-700 km [68, 69]. The relevance of wind for dispersal remains unknown for Lu. shannoni, since high winds can lead to a subsequent reduction in activity [68, 69]. Regardless, wind speed is associated with local weather conditions occurring within defined short periods of time and is not a specific component of larger scale and longer timeframe climate data utilized in ENMs. Further, physical barriers, such as the Rocky Mountain range that spans from western Canada to the southwestern USA, would need to be considered as they may interfere with dispersal. This is particularly relevant since coastal British Columbia is forecasted to be suitable area for Lu. shannoni in the future. However, given there are no known *Lu. shannoni* populations in any proximity to this region and the Rocky Mountains create a large physical barrier, dispersal via natural mechanisms is unlikely [7, 8].

There is a notable lack of continuous surveillance for *Lu. shannoni* across Mexico and the USA. Ecological suitability for the model was inferred using presence-only

data, and given limited surveillance, it is probable that *Lu. shannoni* are present in other regions not incorporated into the model. While increasing the regularization multiplier of the model can reduce the risk of over-fitting the model, it is possible that additional suitable regions for *Lu. shannoni* exist (based on what is known of their ecology). Despite methodological adjustments for the sample size (i.e. setting a regularization multiplier of 1.5, *k*-fold cross-validation), sample size was still relatively small (n=80), which introduces uncertainty and the possibility of model underfitting in some regions while overestimating regions of suitability in others [33, 35, 37, 55–57]. That being said, the high AUC and low mean omissions rate indicate the model is well fitted to the data [33, 35, 37, 55–57].

Regardless, it is important for public health professionals to be aware of current regions throughout Mexico and the USA within the ecological niche of *Lu. shannoni* described here. Due to the vector potential of this sand fly, and its implications for the health of humans, companion animals and livestock, it is recommended that surveillance efforts be concentrated to these highly suitable regions. In the USA and Canada, dogs are imported from *Leishmania*-endemic countries regularly [70–75]. In some instances, infected dogs are imported into Canada [72, 75]. While leishmaniasis cases in humans are rare in the USA and Canada, *Lu. shannoni*'s significance as a potential vector of *L. mexicana* and *L. infantum*, along with VSV in livestock, should encourage surveillance in currently suitable regions to monitor range shifts.

## Conclusions

The ecological niche models generated through this study provide important insights into regions of current and future suitability for *Lu. shannoni* across North America. Surveillance efforts should be directed at these regions to monitor populations and potential range shifts given the public and animal health relevance of this vector. That being said, ecological niche models cannot predict range expansion, and while regions may be identified as being suitable now or in the future for *Lu. shannoni*, future research is needed to explore factors related to dispersal, such as human and animal travel or extreme weather events.

#### Acknowledgements

We would like to thank Dr. Jerome Goddard for providing the coordinates of *Lu. shannoni* capture locations. We would also like to thank Dr. Manisha Kulkarni for providing methodological assistance during MaxEnt model construction.

#### Author contributions

SD conducted data analysis and wrote the main manuscript. KMC provided initial revisions of manuscript and methodological guidance. GKN provided manuscript revisions and methodological guidance. CFP, ALG, JSW provided

revisions on manuscript. All authors reviewed and approved the final version of the manuscript

#### Funding

Research was supported by a Natural Sciences and Engineering Research Council of Canada Discovery Grant (KMC). SD was supported by an Ontario Veterinary College Graduate Scholarship (PhD), and an Ontario Graduate Scholarship.

#### Data availability

Species presence data were obtained from the Global Biodiversity Information Facility (GBIF), the Disease Vectors Database (discontinued) and the National Museum of Natural History (Smithsonian Institution). Additional presence points were obtained from previous surveillance data reported in the literature. Bioclimatic variables were obtained from ClimateNA, and an additional ecoregion layer was obtained from the Commission for Environmental Cooperation.

## Declarations

Ethics approval and consent to participate

Not applicable.

#### **Competing interests**

The authors declare no competing interests.

#### **Consent for publication**

Not applicable.

#### Author details

<sup>1</sup>Department of Population Medicine, Ontario Veterinary College, University of Guelph, Guelph, ON, Canada. <sup>2</sup>Department of Pathology and Microbiology, Faculty of Veterinary Medicine, University of Montreal, Saint-Hyacinthe, QC, Canada. <sup>3</sup>Department of Biology, Trent University, Peterborough, ON, Canada. <sup>4</sup>Department of Pathobiology, Ontario Veterinary College, University of Guelph, Guelph, ON, Canada.

Received: 21 November 2024 Accepted: 28 March 2025 Published online: 25 April 2025

#### References

- Young DG, Duncan MA. Guide to identification and geographic distribution of *Lutzomyia* sand flies in Mexico, the West Indies, Central and South America. Memoirs of the American Entomological Institute Series No. 54. 1994;54. Chicago: Associated Publications; 1994
- Goddard J, McHugh C. New records for the phlebotomine sand fly Lutzomyia shannoni (Dyar) (Diptera: psychodidae) in Mississippi. J Miss Acad Sci. 2005;50:3.
- Haddow AD, Curler G, Moulton JK. New records of *Lutzomyia shannoni* and *Lutzomyia vexator* (Diptera: Psychodidae) in eastern Tennessee. J Vector Ecol. 2008;33:393–6. https://doi.org/10.3376/1081-1710-33.2.393.
- Minter L, Kovacic B, Claborn DM, Lawyer P, Florin D, Brown GC. New state records for *Lutzomyia shannoni* and *Lutzomyia vexator*. J Med Entomol. 2009;46:965–8. https://doi.org/10.1603/033.046.0432.
- González C, Wang O, Strutz SE, González-Salazar C, Sánchez-Cordero V, Sarkar S. Climate change and risk of leishmaniasis in North America: predictions from ecological niche models of vector and reservoir species. PLoS Negl Trop Dis. 2010;4:e585. https://doi.org/10.1371/journal.pntd. 0000585.
- Weng J, Young SL, Gordon DM, Claborn D, Petersen C, Ramalho-Ortigao M. First report of phlebotomine sand flies (Diptera: Psychodidae) in Kansas and Missouri, and a PCR method to distinguish *Lutzomyia shannoni* from *Lutzomyia vexator*. J Med Entomol. 2012;49:1460–5. https://doi.org/ 10.1603/me12105.
- Cecílio P, Cordeiro-Da-Silva A, Oliveira F. Sand flies: basic information on the vectors of leishmaniasis and their interactions with *Leishmania* parasites. Commun Biol. 2022;5:305. https://doi.org/10.1038/ s42003-022-03240-z.

- Maroli M, Feliciangeli M, Bichaud L, Charrel RN, Gradoni L. Phlebotomine sandflies and the spreading of leishmaniases and other diseases of public health concern. Med Vet Entomol. 2013;27:123–47. https://doi.org/10. 1111/j.1365-2915.2012.01034.x.
- Carvalho BM, Rangel EF, Ready PD, Vale MM. Ecological niche modelling predicts southward expansion of *Lutzomyia* (*Nyssomyia*) flaviscutellata (Diptera: Psychodidae: Phlebotominae), vector of *Leishmania* (*Leishmania*) amazonensis in South America, under climate change. PLoS ONE. 2015;10:e0143282. https://doi.org/10.1371/journal.pone.0143282
- DeWinter S, Shahin K, Fernandez-Prada C, Greer AL, Weese JS, Clow KM. Ecological determinants of leishmaniasis vector Lutzomyia spp. a scoping review. Med Vet Entomol. 2024. https://doi.org/10.1111/mve.12741.
- Price DC, Gunther DE, Gaugler R. First collection records of phlebotomine sand flies (Diptera: psychodidae) from New Jersey. J Med Entomol. 2011;48:476–8. https://doi.org/10.1603/me10170.
- Alexander B, Maroli M. Control of phlebotomine sandflies. Med Vet Entomol. 2003;17:1–18. https://doi.org/10.1046/j.1365-2915.2003.00420.x.
- Feliciangeli MD. Natural breeding places of phlebotomine sandflies. Med Vet Entomol. 2004;18:71–80. https://doi.org/10.1111/j.0269-283x.2004. 0487.x.
- Comer JA, Irby WS, Kavanaugh DM. Hosts of *Lutzomyia shannoni* (Diptera: Psychodidae) in relation to vesicular stomatitis virus on Ossabaw Island, Georgia, U.S.A. Med Vet Entomol. 1994;8:325–30. https://doi.org/10. 1111/j.1365-2915.1994.tb00096.x.
- Elias E, Savoy HM, Swanson DA, Cohnstaedt LW, Peters DPC, Derner JD, et al. Landscape dynamics of a vector-borne disease in the western US: how vector–habitat relationships inform disease hotspots. Ecosphere. 2022. https://doi.org/10.1002/ecs2.4267.
- Kaszak I, Planellas M, Dworecka-Kaszak B. Canine leishmaniosis—an emerging disease. Ann Parasitol. 2015;61:69–76.
- 17. Baneth G, Solano-Gallego L. Leishmaniasis. Vet Clin North Am Small Anim Pract. 2022;52:1359–75. https://doi.org/10.1016/j.cvsm.2022.06.012.
- WHO. Leishmaniasis. 2023. https://www.who.int/news-room/fact-sheets/ detail/leishmaniasis. Accessed 28 Aug 2024.
- Gaskin AA, Schantz PM, Jackson JL, Birkenheuer AJ, Tomlinson L, Gramiccia M, et al. Visceral leishmaniasis in a New York foxhound kennel. J Vet Intern Med. 2002;16:34–44. https://doi.org/10.1111/j.1939-1676.2002. tb01604.x.
- Travi BL, Ferro C, Cadena H, Montoya-Lerma J, Adler GH. Canine visceral leishmaniosis: dog infectivity to sand flies from non-endemic areas. Res Vet Sci. 2002;72:83–6.
- Duprey ZH, Steurer FJ, Rooney J, Kirchhoff LV, Jackson JL, Rowton E, et al. Canine visceral leishmaniasis, United States and Canada, 2000–2003. Emerg Infect Dis. 2006;12:440–6. https://doi.org/10.3201/eid1203.050811.
- Gibson-Corley KN, Hostetter JM, Hostetter SJ, Mullin K, Ramer-Tait AE, Boggiatto PM, et al. Disseminated *Leishmania infantum* infection in two sibling foxhounds due to possible vertical transmission. Can Vet J. 2008;49:1005–8.
- Bouattour A, Amri A, Belkhiria JA, Rhim A, Fezaa O, Gantier J, et al. Canine leishmaniosis in Tunisia: growing prevalence, larger zones of infection. PLoS Negl Trop Dis. 2021;15:e0009990. https://doi.org/10.1371/journal. pntd.0009990.
- Morales-Yuste M, Martín-Sánchez J, Corpas-López V. Canine leishmaniasis: update on epidemiology, diagnosis, treatment, and prevention. Vet Sci. 2022;9:387. https://doi.org/10.3390/vetsci9080387.
- Lawyer PG, Young DG, Butler JF, Akin DE. Development of *Leishmania* mexicana in Lutzomyia diabolica and Lutzomyia shannoni (Diptera: *Psychodidae*). J Med Entomol. 1987;24:347–55. https://doi.org/10.1093/ jmedent/24.3.347.
- Lawyer PG, Young DG. Experimental transmission of *Leishmania mexicana* to hamsters by bites of phlebotomine sand flies (Diptera: *Psychodidae*) from the United States. J Med Entomol. 1987;24:458–62. https://doi.org/ 10.1093/jmedent/24.4.458.
- Schaut RG, Robles-Murguia M, Juelsgaard R, Esch KJ, Bartholomay LC, Ramalho-Ortigao M, et al. Vector-borne transmission of *Leishmania infantum* from hounds. United States Emerg Infect Dis. 2015;21:2209–12. https://doi.org/10.3201/eid2112.141167.
- Dergousoff SJ, Galloway TD, Lindsay LR, Curry PS, Chilton NB. Range expansion of *Dermacentor variabilis* and *Dermacentor andersoni* (Acari: lxodidae) near their northern distributional limits. J Med Entomol. 2013;50:510–20. https://doi.org/10.1603/me12193.

- Clow KM, Leighton PA, Ogden NH, Lindsay LR, Michel P, Pearl DL, et al. Northward range expansion of *Ixodes scapularis* evident over a short timescale in Ontario. Canada PLoS ONE. 2017;12:e0189393. https://doi. org/10.1371/journal.pone.0189393.
- Kamal M, Kenawy MA, Rady MH, Khaled AS, Samy AM. Mapping the global potential distributions of two arboviral vectors *Aedes aegypti* and *Ae* albopictus under changing climate. PLoS ONE. 2018;13:e0210122. https://doi.org/10.1371/journal.pone.0210122.
- Khan SU, Ogden NH, Fazil AA, Gachon PH, Dueymes GU, Greer AL, et al. Current and projected distributions of *Aedes aegypti* and *Ae* albopictus in Canada and the U.S. Environ Health Perspect. 2020. https://doi.org/10. 1289/ehp5899.
- Alkishe A, Raghavan RK, Peterson AT. Likely geographic distributional shifts among medically important tick species and tick-associated diseases under climate change in North America: a review. Insects. 2021;12:225. https://doi.org/10.3390/insects12030225.
- Phillips SJ, Anderson RP, Schapire RE. Maximum entropy modeling of species geographic distributions. Ecol Modell. 2006;190:231–59.
- Elith J, Phillips SJ, Hastie T, Dudík M, Chee YE, Yates CJ. A statistical explanation of MaxEnt for ecologists. Divers Distrib. 2010;17:43–57. https://doi. org/10.1111/j.1472-4642.2010.00725.x.
- Merow C, Smith MJ, Silander JA. A practical guide to MaxEnt for modeling species' distributions: what it does, and why inputs and settings matter. Ecography. 2013;36:1058–69. https://doi.org/10.1111/j.1600-0587.2013. 07872.x.
- Escobar LE. Ecological niche modeling: an introduction for veterinarians and epidemiologists. Front Vet Sci. 2020;7:519059. https://doi.org/10. 3389/fvets.2020.519059.
- Morales NS, Fernández IC, Baca-González V. MaxEnt's parameter configuration and small samples: are we paying attention to recommendations? A systematic review. Peer J. 2017. https://doi.org/10.7717/peerj.3093.
- Ferro C, Cárdenas E, Corredor D, Morales A, Munstermann LE. Life cycle and fecundity analysis of *Lutzomyia shannoni* (Dyar) (Diptera: *Psychodidae*). Mem Inst Oswaldo Cruz. 1998;93:195–9. https://doi.org/10.1590/ s0074-02761998000200011.
- Beasley EA, Mahachi KG, Petersen CA. Possibility of *Leishmania* transmission via Lutzomyia spp. sand flies within the USA and implications for human and canine autochthonous infection. Curr Trop Med Rep. 2022;9:160–8. https://doi.org/10.1007/s40475-022-00267-4.
- Adeniran AA, Fernández-Santos NA, Rodríguez-Rojas JJ, Treviño-Garza N, Huerta-Jiménez H, Mis-Ávila PC, et al. Identification of phlebotomine sand flies (Diptera: *Psychodidae*) from leishmaniasis endemic areas in southeastern Mexico using DNA barcoding. Ecol Evol. 2019;9:13543–54. https://doi.org/10.1002/ece3.5811.
- 41. Ibáñez-Bernal S, Muñoz J, Rebollar-Téllez EA, Pech-May A, Marina CF. Phlebotomine sand flies (Diptera: *Psychodidae*) of Chiapas collected near the Guatemala border, with additions to the fauna of Mexico and a new subgenus name. Zootaxa. 2015. https://doi.org/10.11646/zootaxa. 3994.2.1.
- Florin DA, Rebollar-Téllez EA. Divergence of Lutzomyia (Psathyromyia) shannoni (Diptera: Psychodidae: Phlebotominae) is indicated by morphometric and molecular analyses when examined between taxa from the southeastern United States and southern Mexico. J Med Entomol. 2013;50:1324–9. https://doi.org/10.1603/me13085.
- Lozano-Sardaneta YN, Paternina LE, Sánchez-Montes S, Quintero A, Ibáñez-Bernal S, Sánchez-Cordero V, et al. DNA barcoding and fauna of phlebotomine sand flies (Diptera: *Psychodidae: Phlebotominae*) from Los Tuxtlas, Veracruz, Mexico. Acta Trop. 2019;201:105220. https://doi.org/10. 1016/j.actatropica.2019.105220.
- 44. Martínez-Burgos M, Lozano-Sardaneta YN, Rodríguez-Rojas JJ, Gómez-Rivera ÁS, Canto-Mis KL, Flores-Escobar E, et al. Species diversity and detection of pathogens in phlebotomine sand flies collected from forest management areas of Quintana Roo, Mexico. Med Vet Entomol. 2023;37:845–58. https://doi.org/10.1111/mve.12691.
- 45. De Oca-Aguilar AM, Euan-Canul R, Sosa-Bibiano E, López-Ávila K, Rebollar-Téllez E, Palacio-Vargas J, et al. Phlebotomine sand flies in rural Mayan communities of Southern Mexico: the heterogeneity of the ruralscape increases the entomological risk. Acta Trop. 2023;249:107051. https://doi. org/10.1016/j.actatropica.2023.107051.
- Comer JA, Kavanaugh DM, Stallknecht DE, Corn JL. Population dynamics of Lutzomyia shannoni (Diptera: Psychodidae) in relation to the

epizootiology of vesicular stomatitis virus on Ossabaw Island, Georgia. J Med Entomol. 1994;31:850–4. https://doi.org/10.1093/jmedent/31.6.850.

- Comer JA, Kavanaugh DM, Stallknecht DE, Ware GO, Corn JL, Nettles VF. Effect of forest type on the distribution of *Lutzomyia shannoni* (Diptera: *Psychodidae*) and vesicular stomatitis virus on Ossabaw Island, Georgia. J Med Entomol. 1993;30:555–60. https://doi.org/10.1093/jmedent/30.3. 555.
- Comer JA, Corn JL. Funnel trap for the capture of phlebotomine sand flies (Diptera: *Psychodidae*) from tree holes. J Med Entomol. 1991;28:289–92. https://doi.org/10.1093/jmedent/28.2.289.
- Florin DA, Lawyer P, Rowton E, Schultz G, Wilkerson R, Davies SJ, et al. Population dynamics of *Lutzomyia shannoni* (Diptera: *Psychodidae*) at the Patuxent National Wildlife Research Refuge, Maryland. J Am Mosq Control Assoc. 2010;26:337–9. https://doi.org/10.2987/10-6022.1.
- Claborn DM, Rowton ED, Lawyer PG, Brown GC, Keep LW. Species diversity and relative abundance of phlebotomine sand flies (Diptera: *Psychodidae*) on three army installations in the southern United States and susceptibility of a domestic sand fly to infection with Old World *Leishmania major*. Mil Med. 2009;174:1203–8. https://doi.org/10.7205/ milmed-d-00-4309.
- Florin DA, Davies SJ, Olsen C, Lawyer P, Lipnick R, Schultz G, et al. Morphometric and molecular analyses of the sand fly species *Lutzomyia shannoni* (Diptera: *Psychodidae: Phlebotominae*) collected from seven different geographical areas in the southeastern United States. J Med Entomol. 2011;48:154–66. https://doi.org/10.1603/me10199.
- Mann RS, Kaufman PE. The seasonal abundance of phlebotomine sand flies, *Lutzomyia* species in Florida. J Am Mosq Control Assoc. 2010;26:10– 7. https://doi.org/10.2987/09-5901.1.
- Slatculescu AM, Clow KM, McKay R, Talbot B, Logan JJ, Thickstun CR, et al. Species distribution models for the eastern blacklegged tick, lxodes scapularis, and the Lyme disease pathogen, *Borrelia burgdorferi*, in Ontario, Canada. PLoS ONE. 2020;15:e0238126. https://doi.org/10.1371/ journal.pone.0238126.
- Riahi K, Van Vuuren DP, Kriegler E, Edmonds J, O'Neill BC, Fujimori S, et al. The Shared Socioeconomic Pathways and their energy, land use, and greenhouse gas emissions implications: an overview. Glob Environ Change. 2016;42:153–68. https://doi.org/10.1016/j.gloenvcha.2016.05.009
- Fielding AH, Bell JF. A review of methods for the assessment of prediction errors in conservation presence/absence models. Environ Conserv. 1997;24:38–49. https://doi.org/10.1017/S0376892997000088.
- Phillips SJ, Dudík M, Schapire RE. Maxent software for modeling species niches and distributions (version 3.4.4). 2024. http://biodiversityinformat ics.amnh.org/open\_source/maxent/. Accessed 15 Sept 2024.
- Radosavljevic A, Anderson RP. Making better Maxent models of species distributions: complexity, overfitting and evaluation. J Biogeogr. 2013;41:629–43. https://doi.org/10.1111/jbi.12227.
- Oshaghi MA, Ravasan NM, Javadian E, Rassi Y, Sadraei J, Enayati AA, et al. Application of predictive degree day model for field development of sandfly vectors of visceral leishmaniasis in northwest of Iran. J Vector Borne Dis. 2009;46:247–55.
- Davies CE, Reithinger R, Campbell-Lendrum D, Feliciangeli D, Borges RJ, Rodríguez N. The epidemiology and control of leishmaniasis in Andean countries. Cad Saude Publica. 2000;16:925–50. https://doi.org/10.1590/ s0102-311x200000400013.
- Martín ME, Stein M, Willener JA, Kuruc JA, Estallo EL. Landscape effects on the abundance of *Lutzomyia longipalpis* and *Migonemyia migonei* (Diptera: Psychodidae: Phlebotominae) in Corrientes city, northern Argentina. Acta Trop. 2020;210:105576. https://doi.org/10.1016/j.actatropica.2020. 105576.
- 61. Von Stebut E. Leishmaniasis. J German Soc Dermatol. 2015;13:191–200.
- 62. Wiken E, Hirvonen H, Marshall I, Hannah L, Gauthier D, Omernik J, et al. Ecological regions of North America: toward a common perspective. Montreal: Communications and Public Outreach Department, Commission for Environmental Cooperation; 1997.
- 63. Pasos M. Terrestrial ecoregions: level I. Commission for environmental cooperation. http://www.cec.org/north-american-environmental-atlas/ terrestrial-ecoregions-level-i/. Accessed 11 Jul 2024.
- 64. Wang Y, Hogg EH, Price DT, Edwards J, Williamson T. Past and projected future changes in moisture conditions in the Canadian boreal forest. For Chron. 2014;90:678–91. https://doi.org/10.5558/tfc2014-134.

- National Snow and Ice Data Centre. Science of snow. 2025. https://nsidc. org/learn/parts-cryosphere/snow/science-snow#:~:text=For%20snow% 20to%20fall%2C%20moisture,C%20or%2032°F. Accessed 27 Feb 2025.
- National Weather Service. Frost information page. 2025. https://www. weather.gov/dmx/dssfrost. Accessed 27 Feb 2025.
- Flint LE, Flint AL, Thorne JH. Climate change: evaluating your local and regional water resources. Fact sheet. 2015. https://doi.org/10.3133/fs201 43098.
- Sellers RF, Maarouf AR. Trajectory analysis of winds and vesicular stomatitis in North America, 1982–5. Epidemiol Infect. 1990;104:313–28. https:// doi.org/10.1017/s0950268800059495.
- Burgin LE, Gloster J, Sanders C, Mellor PS, Gubbins S, Carpenter S. Investigating incursions of bluetongue virus using a model of long-distance *Culicoides* biting midge dispersal. Transbound Emerg Dis. 2012;60:263–72. https://doi.org/10.1111/j.1865-1682.2012.01345.x.
- Anderson M, Douma D, Kostiuk D, Filejski C, Rusk R, Weese JS, et al. Report of the Canadian National Canine Importation working group. 2016. https://www.canadianveterinarians.net/media/h1cd40v4/report-of-thecanadian-national-canine-importation-working-group.pdf. Accessed 7 Mar 2025.
- Anderson MK, Stull JW, Weese JS. Impact of dog transport on high-risk infectious diseases. Vet Clin North Am Small Anim Pract. 2019;49:615–27. https://doi.org/10.1016/j.cvsm.2019.02.004.
- 72. Gin TE, Lashnits E, Wilson JF, Breitschwerdt EB, Qurollo BA. Demographics and travel history of imported and autochthonous cases of leishmaniosis in dogs in the United States and Canada, 2006 to 2019. J Vet Intern Med. 2021;35:954–64. https://doi.org/10.1111/jvim.16071.
- Julien D, Sargeant JM, Filejski C, Harper SL. Who let the dogs In? An epidemiological study quantifying domestically sourced and imported dogs in Southern Ontario. Canada Zoonoses Public Health. 2021;68:588–600. https://doi.org/10.1111/zph.12847.
- Blackmore J, Gerson H, Clow KM, Anderson MC, Tataryn J. Estimating spatial and temporal trends of dog importation into Canada from 2013 to 2019. Can Vet J. 2023;64:6.
- Wagner VE, Douanne N, Fernandez-Prada C. *Leishmania infantum* infection in a dog imported from Morocco. Can Vet J. 2020;61(9):963– 5. https://europepmc.org/article/MED/32879521

## **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.